

OLIFERCHUK V. P.^{1✉}, FEDOROVYCH D. V.²

¹ National Forestry University of Ukraine,

Ukraine, 79057, Lviv, Gen. Chuprynska str., 103, e-mail: victoriaoliferchuk@gmail.com

² Institute of Cell Biology of Nat. Acad. Sci. of Ukraine,

Ukraine, 79005, Lviv, Drahomanova str., 14/16, e-mail: fedorovych@cellbiol.lviv.ua

✉ victoriaoliferchuk@gmail.com , (066) 464-36-11, (098) 631-36-17

APPLICATION OF MYCORRHIZAL FUNGUS *TUBER MELANOSPORUM* TO STIMULATE THE GROWTH AND DEVELOPMENT OF SOYBEAN AND SPRING BARLEY

Aim. To evaluate the effect of soybean and spring barley seeds treatment with haploid cells of fungus *Tuber melanosporum* IMB F- 100106 on the growth and development of soybean and spring barley. **Methods.** The study was conducted in field conditions on the experimental field of the Department of Agroecology and Biosafety IAP NANU. The level of development of the fungus was evaluated visually and by counting ectomycorrhizal threads. Morphometric and biochemical methods were used to characterize the structure of the crop.

Results. The treatment of soybean and spring barley seeds with *T. melanosporum* promotes productive growth and increases the yield of crops. **Conclusions.** Plant inoculation with *T. melanosporum* promotes productive growth and increases the yield of soybean and barley, promotes the possibility of creating new technologies of organic and regenerative agriculture and allows preserving of the genetic stability of this fungus.

Keywords: mycorrhiza, *Tuber melanosporum*, crop yields, genetic stability.

The importance of mycorrhizal fungi for soil conservation, dynamics of natural ecosystems and sustainable agriculture has been recognized worldwide for more than one century of research on this phenomenon. Mycorrhizal fungi play a significant role in nature: they help plants to better absorb water and nutrients from the soil. This means that a mycorrhizal plant has a better growth and vigor. In addition, the mycorrhizal fungi enhance plants resistance to diseases and drought. Some mycorrhizal fungi produce mushrooms such as *Boletus* and truffles (*Tuber melanosporum*) which are highly valued in the market. *Tuber* species are ectomycorrhizal ascomycetes establishing relationships with different host trees and forming hypogeous fruiting bodies known as truffles. The first experimental proof that *Tuber* spp. have the capacity to establish a symbiotic association with roots of their hosts was

obtained only in the late 1960s [1]. *T. melanosporum* Vittad., the 'black truffle', are in great demand by the food market in many countries because of their special taste and smell, resulting from a blend of hundreds of volatile compounds [2]. The high retail prices can go up to \$ 7000 per kg. Research on these fungi has focused on promoting the cultivation of these fungi to meet increasing worldwide demand and to compensate for the catastrophic decline in their natural production.

T. melanosporum colonizing roots of oak and hazelnut trees is developing typical ectomycorrhizal structures close to the roots of their plant symbionts. These include holm oaks, French oaks, hazelnut, cherry and other deciduous trees. The symbiosis of holm oak saplings and black truffles has been shown to improve photosynthesis and root growth in the plant [3, 4]. In addition, inoculation of *Cistus* and oak seedlings in greenhouses and nurseries with *T. melanosporum* mycelium lead to an increase in plant growth showing that the fungus is a mutualistic symbiont [5, 6]. It has been suggested that *T. melanosporum* may act as a biotroph with an antagonistic nature in the later stages of the interaction, but this trait remains to be experimentally verified. *T. melanosporum* can exist in soils in several forms such as ascospores, germinating ascospores, free-living mycelium, emanating hyphae from mycorrhizas, remainders of ascocarps and ectomycorrhizas, or other forms like mitospores or conidiospores. Studies based on genetic and genomic approaches (i.e. release of the genome sequence) have helped to take a leap toward understanding the life cycle and reproductive modes of these fungi [7]. It was found that the *T. melanosporum* create mycorrhiza with woody species (walnut, hazelnut, fruit trees, deciduous and coniferous species of trees and bushes), as well as raspberries, currants, blueberries [8]. The successful application of inoculation of seeds and soaking root stones of saplings for the improvement of the

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growth and development of various plants, in particular oak and other plants were described [9].

Inoculation with spores is the most common method for producing seedlings colonized with truffle mycorrhizas. However, pure cultures and ectomycorrhizas of *Tuber* have also been used to obtain mycorrhizal plants. Mycelium-based inoculum offers many advantages, such as lower contamination risks and more reliable root colonization. A fed-batch fermentation process of *T. melanosporum* was developed for the efficient production of bioactive mycelia and *Tuber* polysaccharides [10]. Truffle mycelium can be hosted by species, which form ectomycorrhizas like those of the genus *Cistus* [10]. It can be also hosted by orchids without forming a classical ectomycorrhizas structure [11] or hosted by herbaceous plants, which could lodge the mycelium without forming mycorrhizas. The number of *T. melanosporum* genotypes have been recovered in cultivated and natural truffle fields which may have different competitive effects on the other mycobionts [12]. Several other aspects of *T. melanosporum* biology and its interaction with different plants remain unclear, in particular factors involved in ascocarp initiation, the nature of the link between ascocarps and mycorrhizas and atmospheric nitrogen fixation. The below-ground landscape for *T. melanosporum* is characterized by dynamic and complex relationships among fungi, plants, microbes and insects, but the multiple players in this community are not well known, including the fungal community composition [13, 14]. Here, we test the hypothesis that *T. melanosporum*, can develop on the roots of leguminous and cereal plants and influence their growth and development.

Materials and methods

Two species of plant: spring barley, hybride "Patricia" and soybean variety "Mercury" – were used as a research objects. For inoculation of plants we used preparation "Micovital" active ingredient of which is *Tuber melanosporum* IMB F-100106 strain isolated from the natural environment, namely from the fruit body of the Perigorsky truffle. The applied agent contained $7 \times 10^9 \pm 5\%$ of vegetative cells *Tuber melanosporum* VS 1223 per dmi. The effect of mycorrhization was studied in field conditions on the experimental field of the Department of Agroecology and Biosafety IAP NAN (Bilyi Rukav village, Khmelnytsky region, Vinnytsia region) – Forest-steppe of Ukraine. The area of the experimental plot was 25 m².

The solutions of the agent acted in a range of concentrations: for the processing of soya seeds – 1 l/t, for the treatment of barley seeds – 0.5 l/t for 30 minutes. Standard preparation.

MikePro PS 3, p. (247 g /ha of seed quantity) were used as a control. The intensity of the growth processes of barley was estimated at the height of the stem when the tube was emitted (Ls-1), the number of productive stems, plant density, growth vigor, and grain productivity components were determined after harvesting, among them: weight of 1000 grains, 1000W, protein and starch content in grain, % of grain content. The intensity of soybean growth processes was evaluated in a 3–5 leaf stage, determining the degree of formation and development of the notation apparatus. After harvesting, the number and weight of tubers, the number of beans and seeds in beans, weight of 1000 seeds, yield, oil content and protein content were determined.

Results and discussion

Pre-sowing treatment of soybean seeds with *T. melanosporum* positively influenced the formation and development of the plant's nodular structure and, as a result, contributed to its active functioning (Fig. 1, table 1).

The number of nodules increased by 12.4% and their weight by 14.4% compared with the values of the control variant. During the flowering phase, in which actually occurs the most active process of biological fixation of nitrogen, all the nodules were bright red in color, which is evidence of the activity of the nitrogenous system. In addition a 9.5% increase in the number of plants per m² was noted, compared to the control variant. The indicated parameters for the test strain exceeded the parameters for the standard preparation MikePro PS 3, p. (247 g /ha of seed quantity) the number of nodules – by 7.0%, the weight of nodules – by 6.2%, the number of plants per m² – by 4.5%. The use of the test *T. melanosporum* VS 1223 fungus strain caused the improvement of the elements of the structure of soybean yield (Table 2). The application of the test preparation for pre-sowing seed treatment lead to a 15.0% increase in the number of beans per plant, the number of beans per m² increased by 26.0%, and the weight of 1,000 seeds – by 4.5%, compared to the control variant indexes.

In all the experiments using the *T. melanosporum* the obtained values were higher than in the variant with the standard product MikePro PS 3, p. The exceedance of the test agent over the standard

product MikePro PS 3, p. was 3.1%. An important indicator that determines the biological value of soybeans is the protein content of seeds that is characterized by unique properties, and its amino acid composition is equivalent to animal proteins and vegetable fat, which is responsible for the energy value of soy as a raw material. Characteristics of quantity and quality of soybean under the application of *T. melanosporum* are given in Table 3.

The application of test strain of the *T. melanosporum* resulted in increase of yield of soybean,

oil and protein content in seeds. The values of these indices were higher than with the standard MikePro PS 3,p. preparation.

The effectiveness of the biological action *T. melanosporum* was tested on the spring barley Barley seeds were treated with *T. melanosporum* in the 7×10^9 titre of cells per cm^3 of the medium at the rate of 1 L per ton of seed. Obtained results are presented on fig. 2 and table 4.



Fig. 1. The root of soybean treated with *T. melanosporum* (2) and without treatment (1).

Table 1. The influence of inoculation soybean seeds with *T. melanosporum* in the 7×10^9 titre of cells per cm^3 of the medium on the growth and development of soybean

| Variant | Number of nodules pcs/plant | Weight of nodules, g/plant | Number of plants pcs/ m^2 |
|--|--------------------------------|-------------------------------|---------------------------------------|
| Control (without treatment) | 29.7 ± 0.62 | 1.04 ± 0.03 | 60.9 ± 1.08 |
| Standard preparation MikePro PS 3, p. | 31.2 ± 0.75 | 1.12 ± 0.04 | 63.8 ± 1.1 |
| <i>T. melanosporum</i> | 33.4 ± 0.51 | 1.19 ± 0.02 | 66.7 ± 1.08 |

Note. pcs – pieces.

Table 2. The influence of inoculation soybean seeds with *T. melanosporum* in the 7×10^9 titre of cells per cm^3 of the medium on the elements of soybean yield structure

| Variant | Number of beans, pcs/plant | Number of beans, pcs/ m^2 | Number of seeds, pcs/bean | Weight of 1,000 seeds, g |
|--|-------------------------------|---------------------------------------|---------------------------------|-----------------------------|
| Control (without treatment) | 41.2 ± 1.7 | $2,509.1 \pm 105.7$ | 3.0 ± 0.04 | 232.1 ± 1.5 |
| Standard preparation MikePro PS 3, p. | 44.7 ± 1.4 | $2,851.9 \pm 106.2$ | 3.0 ± 0.05 | 239.4 ± 1.1 |
| <i>T. melanosporum</i> | 47.4 ± 1.4 | $3,161.6 \pm 107.8$ | 3.0 ± 0.03 | 242.6 ± 1.1 |

Table 3. The influence of inoculation soybean seeds with *T. melanosporum* in the 7×10^9 titre of cells per cm³ of the medium on yield and quality of soybean

| Variant | Yield, ton/ha | Increase, % of control | Oil con- tent, % | Protein content, % |
|-----------------------------|------------------|---------------------------|---------------------|-----------------------|
| Control (without treatment) | 2.64 ± 0.12 | — | 21.5 ± 0.7 | 38.6 ± 0.7 |
| MikePro PS 3, p. | 2.92 ± 0.15 | 10.6 ± 0.05 | 22.1 ± 0.6 | 39.9 ± 0.6 |
| <i>T. melanosporum</i> | 3.01 ± 0.10 | 14.0 ± 0.02 | 23.2 ± 0.9 | 41.5 ± 0.7 |



Fig. 2. The root of spring barley treated with *T. melanosporum* (1), MikePro PS 3, p (2) and without treatment (3).

Table 4. Influence of inoculation barley seeds with *T. melanosporum* on the growth and development of spring barley

| Variant | Number of stems, pcs / m ² | | Height of plants, see | Plant density, psc / m ² | Growth force, % |
|---------------------------------------|---------------------------------------|-----------------|-----------------------|-------------------------------------|-----------------|
| | of all | Productive | | | |
| Control (without treatment) | 560.5 ± 6.4 | 526.3 ± 3.1 | 62.5 ± 1.8 | 365.8 ± 1.3 | 77.2 |
| Standard preparation MikePro PS 3, p. | 589.7 ± 6.1 | 569.4 ± 2.5 | 67.3 ± 1.7 | 391.3 ± 1.2 | 85.6 |
| <i>T. melanosporum</i> | 601.4 ± 5.8 | 588.7 ± 2.7 | 71.8 ± 1.6 | 408.7 ± 1.5 | 89.5 |

Notes: pcs – pieces; see – centimeters.

The use of the test agent showed positive results regarding the growth and development of spring barley (table 4). The morphometric indices of cultivated plants increased: plant height – by 14.9%, density – by 11.7%, growth rate – by 12.3%, compared to the control variant. Also, the total number of stems per 1 m² was increased by 7.3%, and the number of productive stems per 1 m² by 11.8%, relative to control.

Growth characteristics of spring barley increased with application of the study agent also in comparison with the reference product MikePro PS 3, p. (247 g per hectare of seed rate): the total number of stems per 1 m² increased by 2.0%, the number of productive stems increased by 3. 4%,

plant height – by 6.7%, plant density – by 4.4%, growth rate – by 3.9%, which is evidence of the effectiveness of *T. melanosporum* and justifies the feasibility of its use for pre-sowing treatment of spring barley seeds.

Important indicators of grain crops are yield and quality of grain. The treatment of barley seeds by *T. melanosporum* positively influenced the yield and quality characteristics of the grain (Table.5). As a result of pre-seed treatment, the yield of barley grain increased by 13.2%, compared with the control variant and by 4.7%, compared to the reference product. The weights of 1000 grains increased by 9.1% and 3.1%, respectively.

Table 5. Yield and quality of barley grain treated with *T. melanosporum*

| Variant | Yield, c/ha | Increase, % | Weight of 1000 seeds, g | Contents % | | Filamentality of grain |
|---------------------------------------|----------------|----------------|-------------------------------|------------|----------|---------------------------|
| | | | | Protein | Starch | |
| Control (without treatment) | 53.1±0.5 | — | 39.6±0.1 | 10.2±01 | 58.6±01 | 9.6±0.2 |
| Standard preparation MikePro PS 3, p. | 57.4±0.1 | 8.1±0.1 | 41.9±0.4 | 11.0±0.3 | 61.2±0.1 | 9.4±0.3 |
| <i>T. melanosporum</i> | 60.1±0.2 | 13.2±0.1 | 43.2±0.2 | 11.7±0.1 | 64.1±0.1 | 9.1±0.5 |

Qualitative characteristics of barley grain of yarrow determine the content of its composition of starch and protein. As a result, starch content increased by 5.5%, compared to the control and by 2.9% compared to the reference product. The content of protein in the grain of barley has a similar tendency to increase by 1.5% and 0.7% respectively. The straw content of grains of barley of the fresh straw of Patricia is 0.5%, compared to the control variant and 0.3%, compared with the reference product

Conclusions

Inoculation of soybean and spring barley with *T. melanosporum* promotes productive growth and increases the yield of crops, promotes the possibility of creating new technologies of organic and regenerative agriculture. The above observations obviously need further work to provide more definitive answers to the functional aspects of the association of *T. melanosporum* mycelium with the non-host plants. However, already at this stage, it appears clearly that direct *T. melanosporum* interactions with non-host plants do exist, and that improved knowledge on these interactions can be used to promote new technologies of organic and regenerative agriculture and allows to preserve the genetic stability of this fungi species.

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ОЛІФЕРЧУК В. П.¹, ФЕДОРОВИЧ Д. В.²

¹ Національний лісотехнічний університет України,
Україна, 79057, м. Львів, вул. Ген. Чупринки, 103, e-mail: victorijaoliferchuk@gmail.com

² Інститут біології клітини НАН України,
Україна 79005, м. Львів, вул. Драгоманова, 14/16, e-mail: fedorovych@cellbiol.lviv.ua

ЗАСТОСУВАННЯ МІКОРИЗАЛЬНОГО ГРИБА *TUBER MELANOSPORUM* ДЛЯ СТИМУЛЯЦІЇ РОСТУ І РОЗВИТКУ СОЇ ТА ЯРОГО ЯЧМЕНЮ

Мета. Оцінити вплив обробки насіння сої та ярого ячменю гаплоїдними клітинами гриба *Tuber melanosporum* IMB F-100106 на ріст і розвиток сої та ярого ячменю. **Методи.** Дослідження проводили у польових умовах на дослідному полі відділу агроекології і біобезпеки ІАП НАН України. Рівень розвитку гриба визначали візуально і кількісно шляхом підрахунку ектомікоризних ниток. Морфометричні та біохімічні методи застосовували для характеристики структури врожаю. **Результати.** Обробка насіння сої та ярого ячменю гаплоїдними клітинами *T. melanosporum* сприяє продуктивному росту і збільшує врожайність сої та ярого ячменю. **Висновки.** Інокуляція рослин *T. melanosporum* покращує ріст і підвищує врожайність сої і ячменю, сприяє можливості створення нових технологій органічного та регенеративного землеробства та дозволяє зберегти генетичну стабільність цього гриба.

Ключові слова: мікориза, *Tuber melanosporum*, врожайність, генетична стабільність.